



Anti-EBV-CA ELISA (IgG)



- **Highly specific and sensitive test for the detection of Epstein-Barr virus antibodies**
- **Ideally suited for the determination of the EBV immune status (seronegativity/seropositivity)**
- **More tests available for avidity determination and CSF diagnostics**

Technical data

| | |
|------------------------|--|
| Antigen | Purified Epstein-Barr virus capsid antigens; antigen source: inactivated cell lysate of human B cells infected with Epstein-Barr virus of strain P3HR1 |
| Calibration | Quantitative, in relative units per ml (RU/ml) Calibration serum 1: 200 RU/ml Calibration serum 2: 20 RU/ml Calibration serum 3: 2 RU/ml Recommended upper threshold of the reference range for non-infected individuals (cut-off): 20 RU/ml |
| Sample dilution | Serum or plasma, 1:101 in sample buffer |
| Reagents | Ready for use, with the exception of the wash buffer (10x); colour-coded solutions, in most cases exchangeable with those in other EUROIMMUN ELISA kits |
| Test procedure | 30 min / 30 min / 15 min, room temperature, fully automatable |
| Measurement | 450 nm, reference wavelength between 620 nm and 650 nm |
| Test kit format | 96 break-off wells, kit includes all necessary reagents |
| Order number | EI 2791-9601 G |

Clinical significance

EBV (Epstein-Barr virus) and herpes simplex virus types 1 and 2 belong to the most ubiquitous human-pathogenic herpes viruses in adults. The virus is the causative agent of infectious mononucleosis (glandular fever), a febrile disease usually accompanied by pharyngitis and lymphadenopathy, frequently by hepatosplenomegaly and more rarely by exanthema. EBV infections are also found in connection with Burkitt's lymphoma and nasopharyngeal carcinoma. The clinical picture of EBV infection can be diverse. The symptoms are unspecific and often overlap with those of other diseases. EBV infection should be differentiated diagnostically from infections with CMV, Toxoplasmosa, Streptococcus, parvovirus B19 and HIV.

Diagnostic application

Since direct detection of EBV is often difficult, serological tests are routinely used for diagnosing EBV infections. The immune response after infection is characterised by the development of antibodies against the EBV capsid antigen (EBV-CA), the EBV nuclear antigens (EBNA-1 to EBNA-6) and the EBV early antigens (EBV-EA). In over 90% of cases an acute EBV infection can be characterised serologically by the detection of anti-EBV-CA IgM and an increase in titer of anti-EBV-CA IgG using ELISA. An at least twofold increase in the anti-EBV-CA IgG titer and the absence of antibodies against EBNA-1 is characteristic for the early phase of an acute EBV infection. Serologically challenging constellations can be clarified by measuring the avidity of the anti-EBV-CA IgG antibodies (EI 2791-9601-1 G). EBV infections of the central nervous system can be diagnosed by determining the anti-EBV-CA antibodies of class IgG in the cerebrospinal fluid (EI 2791-9601-L G).



Linearity

The linearity of the Anti-EBV-CA ELISA (IgG) was determined by performing four serial dilutions of different serum samples. The linear regression R^2 was > 0.95 for all samples. The Anti-EBV-CA ELISA (IgG) is linear in the investigated concentration range (4–141 RU/ml).

Reference range

Levels of anti-EBV-CA antibodies (IgG) were analysed in a group of 500 healthy blood donors using the EUROIMMUN ELISA. With a cut-off value of 20 RU/ml, 93.4% of the blood donors were anti-EBV-CA positive (IgG), in agreement with the known infection level in adults.

Reproducibility

The reproducibility was investigated by determining the intra- and inter-assay coefficients of variation using three sera. The intra-assay CVs are based on 20 determinations and the inter-assay CVs on four determinations performed in six different test runs.

| Serum | Intra-assay variation, n=20 | | Inter-assay variation, n=4x6 | |
|-------|-----------------------------|--------|------------------------------|--------|
| | Mean value (RU/ml) | CV (%) | Mean value (RU/ml) | CV (%) |
| 1 | 47 | 7.4 | 47 | 8.2 |
| 2 | 90 | 5.8 | 90 | 3.2 |
| 3 | 93 | 4.2 | 93 | 5.4 |

Specificity and sensitivity

In a panel of 175 clinically and serologically precharacterised patient samples (quality assessments by INSTAND, Germany / Labquality, Finland) were investigated using the EUROIMMUN ELISA. The specificity and sensitivity were each 100%, excluding borderline sera.

| n=175 | | INSTAND/Labquality | | |
|--------------------------------------|------------|--------------------|------------|----------|
| | | positive | borderline | negative |
| EUROIMMUN Anti-EBV-CA ELISA (IgG) | positive | 145 | 0 | 0 |
| | borderline | 3 | 1 | 0 |
| | negative | 0 | 0 | 26 |

Prevalence

Sera from children, pregnant women and healthy blood donors were investigated for IgG and IgM antibodies using the EUROIMMUN Anti-EBV-CA ELISA. The prevalences corresponded to the data found in literature (e.g. Bauer, G: Rationale und rationelle Epstein-Barr-Virus-Diagnostik, Clin Lab, 1995).

| Panel | n | Positive results EUROIMMUN Anti-EBV-CA ELISA | | |
|---------------------------------|-----|---|------|----------|
| | | IgG | IgM | IgG, IgM |
| Healthy children ≤ 3 years | 25 | 20.0% | 0.0% | 20.0% |
| Healthy children 4–10 years | 63 | 49.2% | 1.6% | 49.2% |
| Pregnant women | 100 | 98.0% | 0.0% | 98.0% |
| Healthy blood donors | 500 | 93.4% | 1.0% | 93.6% |

Literature

1. Maeda E, Akahane M, Kiryu S, Kato N, Yoshikawa T, Hayashi N, Aoki S, Minami M, Uozaki H, Fukayama M, Ohtomo K. Spectrum of Epstein-Barr virus-related diseases: a pictorial review. J Clin Virol 27 (2009) 4-19.
2. EUROIMMUN AG. Stöcker W, Schlumberger W. All entries on autoimmune diagnostics and laboratory diagnostics in infectious diseases. In: Gressner A, Arndt T (Eds.) Lexikon der Medizinischen Laboratoriumsdiagnostik. 2nd ed., Springer Medizin Verlag, Heidelberg (2012)
3. Balfour Jr H, Dunmire S, Hogquist A. Infectious mononucleosis. Clin Transl Immunology 2015 Feb; 4(2): e33