Anti-Zika Virus ELISA (IgAM)

- Combined detection of IgA and IgM antibodies against Zika virus to support diagnosis of acute infections
- Cross reactions are virtually excluded due to the use of virus-specific NS1 antigen
- Discrimination from other symptomatically similar viral infections (e.g. dengue or chikungunya)

### Technical data

**Antigen**
Recombinant non-structural protein (NS1) of Zika virus

**Calibration**
Semi-quantitative; calculation of a ratio from the extinction of the sample and the extinction of the calibrator

**Result interpretation**
EUROIMMUN recommends interpreting results as follows:
- Ratio < 0.8: negative
- Ratio ≥ 0.8 to < 1.1: borderline
- Ratio ≥ 1.1: positive

**Sample dilution**
Serum or plasma, 1:101 in sample buffer

**Reagents**
Ready for use, with the exception of the wash buffer (10x); colour-coded solutions, in most cases exchangeable with those in other EUROIMMUN ELISA kits

**Test procedure**
60 min (37°C) / 30 min / 15 min, room temperature, fully automated

**Measurement**
450 nm, reference wavelength between 620 nm and 650 nm

**Test kit format**
96 break-off wells; kit includes all necessary reagents

**Order number**
El 2668-9601 Q

**Related products**
El 2668-9601 A, G or M: Anti-Zika Virus ELISA (IgA, IgG or IgM)

### Clinical significance

Zika virus (ZIKV) is an arbovirus of the Flaviviridae family. The virus is generally not transmitted between humans. In individual cases, however, transmission via sexual intercourse has been reported. The virus is transmitted through bites of mosquitoes of the genus Aedes. The virus was first observed in African countries. In recent years there have been major outbreaks in tropical and subtropical regions in Asia and on Pacific islands. Recently an increasing number of infections has been observed in Latin America. In most cases the disease course is mild. The symptoms are near-to identical to those of dengue or chikungunya virus infections. After an incubation time of five to ten days a flu-like illness develops with fever, rash, arthralgia, myalgia, headache and conjunctivitis. During the Zika outbreaks 2014 in Polynesia and 2015/2016 in Latin America, a significant increase in neurological diseases such as Guillain-Barré syndrome was registered. Especially in Brazil, an exceptionally high number of babies were born with microcephaly, a cranio- cephalic malformation. The link between the presence of a ZIKV infection and the occurrence of neurological diseases or foetal malformations is considered as virtually proven. There is no specific treatment for ZIKV infection. Protection from mosquito bites serves as a preventative measure. Since the detection of the virus or its components is only possible during the viraemic phase (up to one week after infection) and the relatively mild disease course often causes patients to seek medical advice at an advanced stage of disease, serological investigation is of major importance. Specific antibodies can be detected several days after the onset of symptoms.

### Diagnostic application

The EUROIMMUN Anti-Zika Virus ELISA (IgAM) is a screening test for the serodiagnosis of acute ZIKV infections. Due to the use of virus-specific NS1 antigen, cross reactions can be virtually excluded. Thus, ZIKV infections can be discriminated from infections with other viruses such as dengue and chikungunya, which cause similar symptoms and are endemic in the same regions. To support the specific IgAM detection, IgG should also be determined. Seroconversion or a significant increase in the IgG antibody titer also indicates an acute infection.
Reference range

The levels of anti-ZIKV antibodies (IgAM) were analysed with this EUROIMMUN ELISA in a panel of 500 healthy blood donors (origin: Germany). With a cut-off of ratio 1.0, no anti-ZIKV antibodies (IgAM) were detected.

Reproducibility

The reproducibility of the test was investigated by determining the intra- and inter-assay coefficients of variation using six samples. The intra-assay CVs are based on 20 determinations and the inter-assay CVs on two determinations performed in 14 different test runs.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Intra-assay variation, n=20</th>
<th>Inter-assay variation, n=2 × 14</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean value (ratio)</td>
<td>CV (%)</td>
</tr>
<tr>
<td>1</td>
<td>0.2</td>
<td>3.1</td>
</tr>
<tr>
<td>2</td>
<td>0.3</td>
<td>2.0</td>
</tr>
<tr>
<td>3</td>
<td>0.9</td>
<td>6.0</td>
</tr>
<tr>
<td>4</td>
<td>0.9</td>
<td>5.5</td>
</tr>
<tr>
<td>5</td>
<td>2.3</td>
<td>2.5</td>
</tr>
<tr>
<td>6</td>
<td>2.7</td>
<td>5.5</td>
</tr>
</tbody>
</table>

Sensitivity and specificity

Samples from 31 patients (origin: Dominican Republic), whose first samples had been classified as positive by ZIKV RT-PCR, were investigated with the EUROIMMUN Anti-Zika Virus ELISA (IgAM). The samples were taken >5 days after onset of symptoms.

The control panel comprised serum samples from 40 patients (origin: Vietnam), in whom a dengue virus infection could be confirmed by direct pathogen detection and serology (day 3–7 after onset of symptoms).

Of the 31 patient samples, all were positive for combined specific IgA and IgM in the EUROIMMUN Anti-Zika Virus ELISA (IgAM). Anti-ZIKV NS1 IgM was detected in 29% (9/31) of the samples. In the control panel none of the sera reacted in the Anti-Zika Virus ELISA (IgM), while two samples (5.0%) reacted in the Anti-Zika Virus ELISA (IgAM).

Cross reactivity

The recombinant NS1 protein of ZIKV has proven to be a highly specific target structure for the immune response in internal and external studies. Despite significant homologies within the flavivirus genus, cross reactions can be virtually excluded when using the NS1 protein. For investigation of the cross reactivity, sera from patients with dengue virus infections, from patients after TBE or yellow fever vaccination and from patients with HCV or WNV infections were analysed. Only one patient from the panel showed a positive result with the Anti-Zika Virus ELISA (IgAM).

Note: It must be taken into account that double infections are possible, especially in endemic regions, or that an infection with another flavivirus may have occurred at an earlier time point. In this case, positive results are not caused by a cross reactivity of the corresponding antibodies. Since interference in samples with acute Plasmodium spp. infections cannot be excluded, malaria should always be taken into account in differential diagnosis.

Literature